

LE MACROALGHE



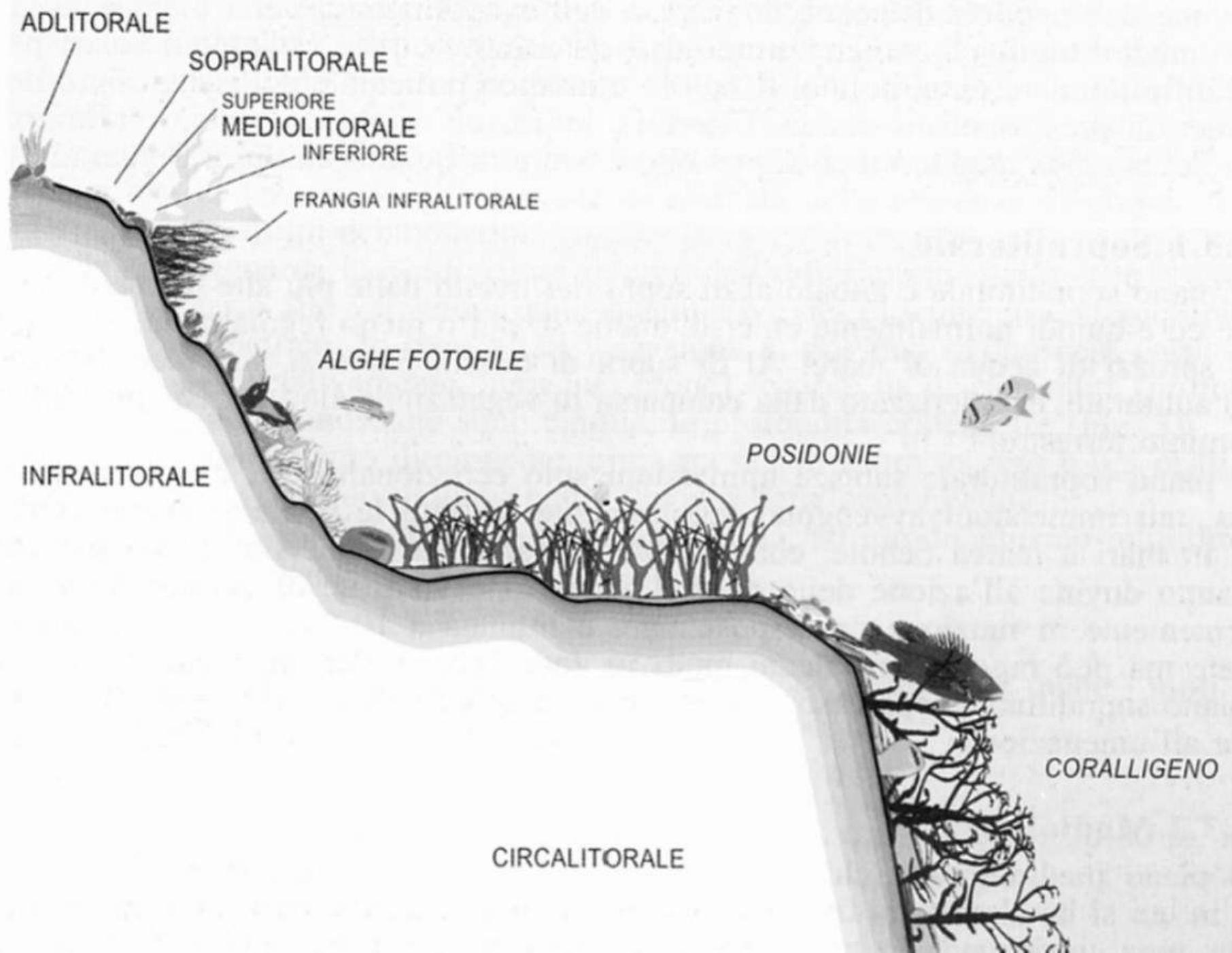
Prof.ssa Giulia Ceccherelli

Una pianta



Un'alga





Caratteristiche

- morfologiche
- tassonomiche
- adattamenti
- utilizzo
- ruolo ecologico
- alge minacciose
- minacce per le alge

Gruppi morfo-funzionali	Morfologia esterna	Anatomia interna	'Texture'	Appetibilità	Durezza	Generi rappresentativi
Appiattite	Tubulari sottili o come un foglio	Non corticate, spessore di una cellula o più	Soffice	Alta	Bassa	<i>Ulva, Enteromorpha, Porphyra</i>
Filamentose	Delicatamente ramificate (filament.)	Uniseriate, multiseriate leggermente corticate	Soffice	Alta	Bassa	<i>Caetomorpha, Cladophora, Ceramium</i>
Ramificate	Fortemente ramificate, erette	Parenchimatose o pseudoparenchimatose	'Fleshy'	Moderata	Bassa	<i>Gigartina, Chondrus, Agardhiella</i>
'a pelle spessa'	Fronde spesse e ramificate	Parenchimatose o pseudoparenchimatose	Simile a pelle fino a gommosa	Bassa	Alta	<i>Fucus, Laminaria, Sargassum, Padina</i>
Calcaree	Articolate, calcarizzate, erette	Segmenti calcificati, giunture flessibili	Simile a pietra	Bassa	Molto alta	<i>Halimeda, Corallina</i>
Crostate	Prostrate, incrostanti	Calcificate o non, file di cellule compatte	Simile a pietra ma dura	Molto Bassa	Molto alta	<i>Hildenbrandia, Ralfsia</i>

Figure 9.16. Algal functional groups as related to molluscan grazer activity. (a) The functional groups; grazing difficulty refers only to structural toughness of the algae and does not take into account the additional difficulty of grazing algal groups 3, 4, and 6 due to their size refuge. (b) Relative importance of each algal group in the diets of 106 species of herbivorous mollusk. (From Steneck & Watling 1982, with permission of Springer-Verlag)

Functional group	Representatives	Morphology	Anatomy	Grazing difficulty
1. Microalgae	Diatoms blue-greens	[50 µm]	[50 µm]	
2. Filamentous algae	<i>Cladophora</i> <i>Ectocarpus</i> <i>Acrochaetium</i>	[1 mm] [300 µm]	[50 µm]	
3. Foliose algae	<i>Ulva</i> <i>Porphyra</i>	[5 cm]	[40 µm]	
4. Corticated macrophytes	<i>Bryothamnium</i> <i>Chondria</i> <i>Acanthophora</i>	[5 cm]	[250 µm]	
5. Leathery macrophytes	<i>Laminaria</i> <i>Fucus</i> Noncalcareous crust	[50 cm] [10 cm]	[200 µm]	
6. Articulated calcareous algae	<i>Halimeda</i> <i>Corallina</i>	[1 cm]	[500 µm]	
7. Crustose coralline algae	Crustose corallines	[3 cm]	[50 µm]	

(a)

Appiattite



Ulva sp.





Chaetomorpha sp.

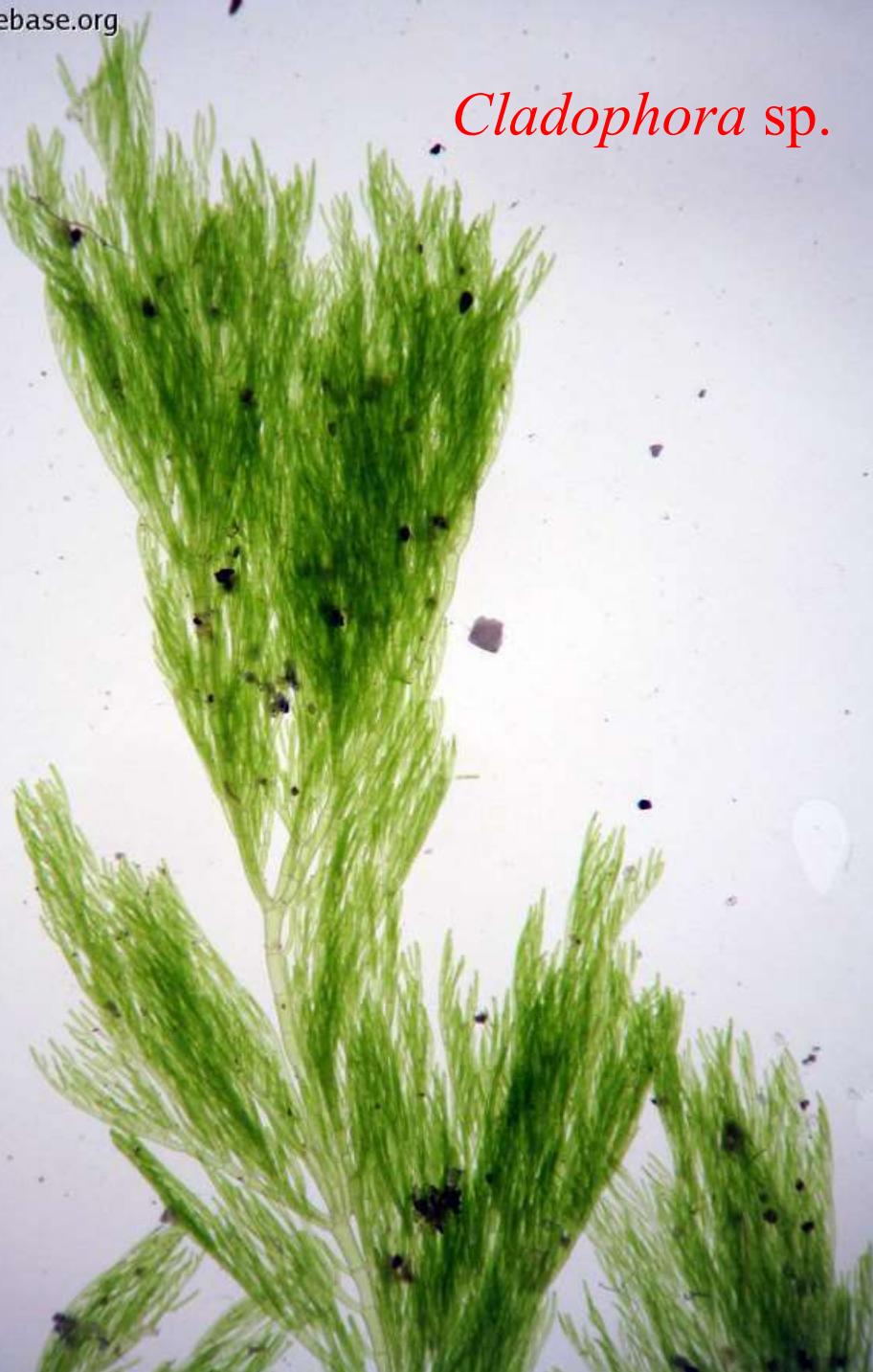
Filamentose



Cladophora sp.

Cladophora sp.

Filamentose



Chondrus crispus

www.algaebase.org

Ramificate

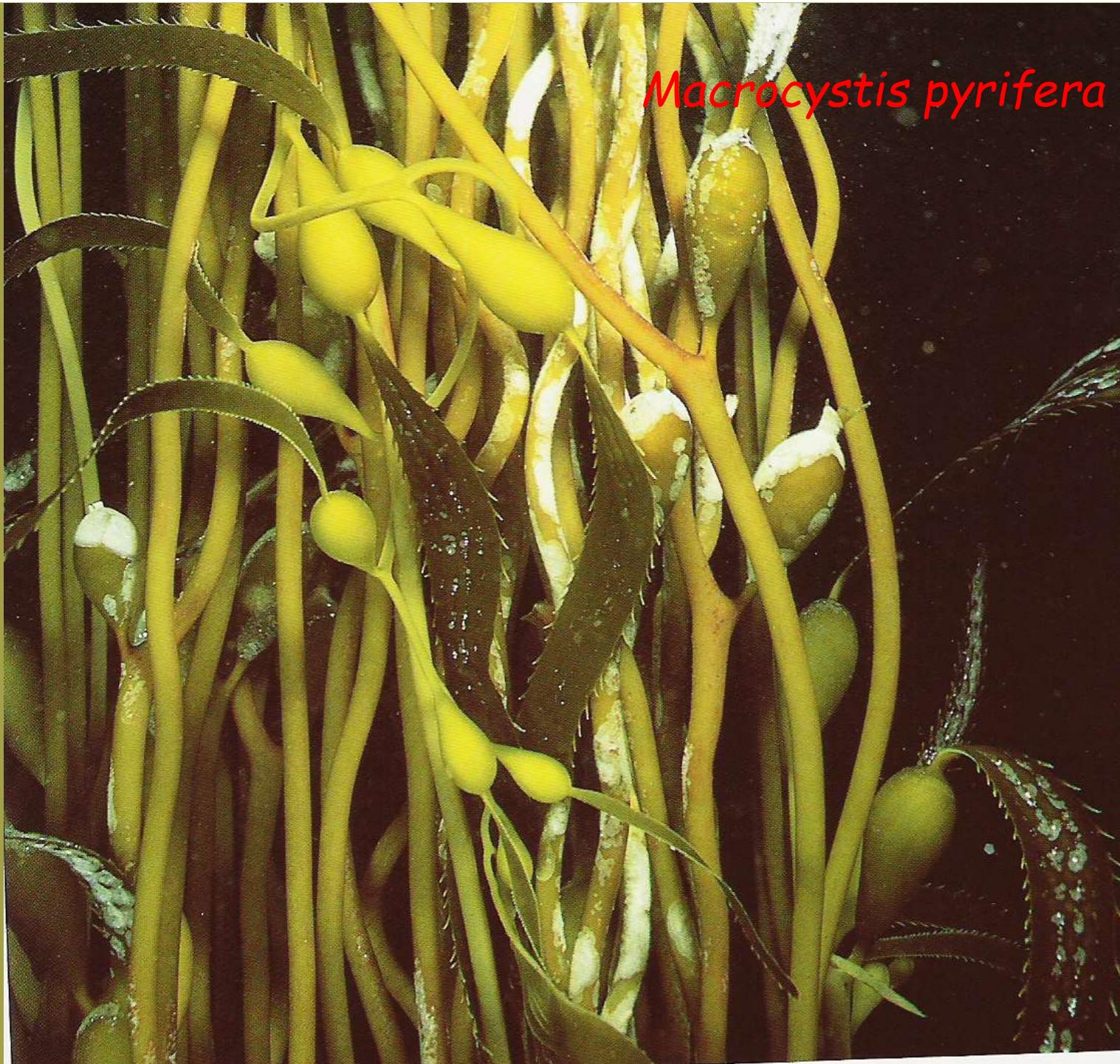


Fucus vesiculosus

A pelle spessa



Macrocystis pyrifera





Corallina sp.

Calcaree

Hildenbrandia sp.

Crostose

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Hildenbrandia sp.

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Cryptogamae: I-XIII, XIV.1-4, XV-XX; Phanerogamae: XIV.5-9; Thallophyta: II-XII, XV-XX; Algae: II-XII; Fungi: Gymnospermae: XIV.5-8; Angiospermae: XIV.9; XI-XXVI; Parazoa (= Porifera): XXVII; and (Eu)metazoa:

Le macroalghe

Regnum	Divisio (phylum) (Division)	Classis (Class)	Fig.
Phyla of Archaeabacteria			
Alge verdi	I. Other phyla of the Eubacteria		
Alge brune	II. Cyanophyta (= Cyano-bacteria)	1. Cyanophyceae	6
Alge rosse	III. Prochlorophyta (= Chloroxybacteria)	1. Prochlorophyceae	7
	IV. Glaucophyta	1. Glaucophyceae	8
	V. Rhodophyta	1. Bangiophyceae 2. Florideophyceae	10 11
c. EUKARYOTA 'Plants' ('Plantae')	VI. Heterokontophyta	1. Chrysophyceae 2. Parmophyceae 3. Sarcinochrysidophyceae 4. Xanthophyceae 5. Eustigmatophyceae 6. Bacillariophyceae 7. Raphidophyceae 8. Dictyochophyceae 9. Phaeophyceae 10. Oomycetes	5 6 7,8 9
	VII. Haptophyta	1. Haptophyceae	2
	VIII. Cryptophyta	1. Cryptophyceae	3
	IX. Dinophyta	1. Dinophyceae	1
	X. Euglenophyta	1. Euglenophyceae	4
	XI. Chlorarachniophyta	1. Chlorarachniophyceae	
	XII. Chlorophyta	1. Prasinophyceae 2. Chlorophyceae 3. Ulvophyceae 4. Cladophorophyceae 5. Bryopsidophyceae 6. Zygnematophyceae 7. Trentepohliophyceae 8. Klebsormidiophyceae 9. Charophyceae	12 14 16 13 15

Alge rosse

Alge brune

Alge verdi

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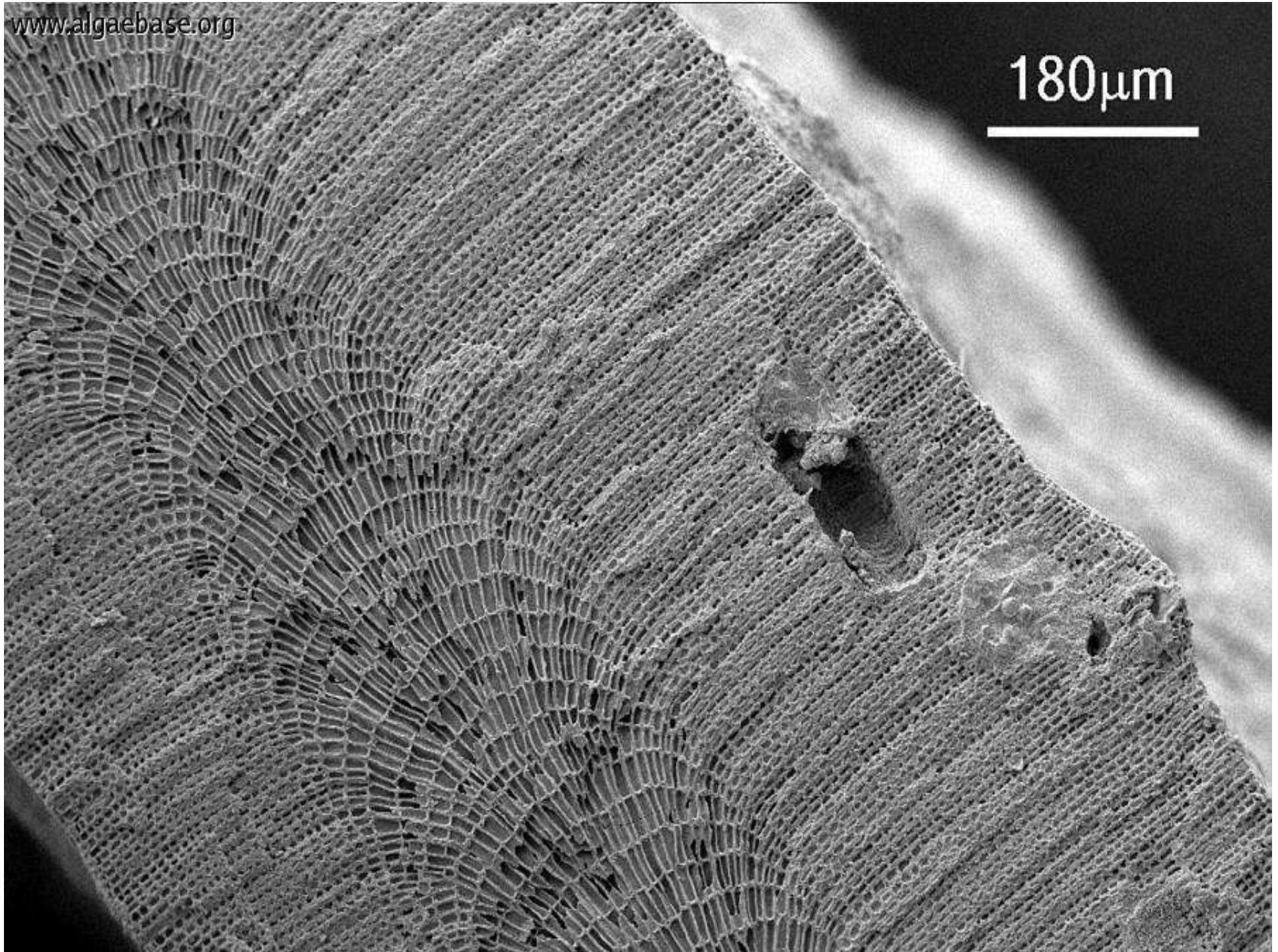
Lithophyllum frondosum

Adattamento morfologico

Lithophyllum dentatum



180 μ m





Lithophyllum frondosum



Lithophyllum ispidum



In condizioni ottimali forma marciapiedi organogeni (trottoir) su substrati suborizzontali e cornici su substrati subverticali. In condizioni meno favorevoli forma cuscini mammellonati sparsi e croste squamate.





Adattamento fotocromatico

La distribuzione dei pigmenti
nei vari gruppi di alghe

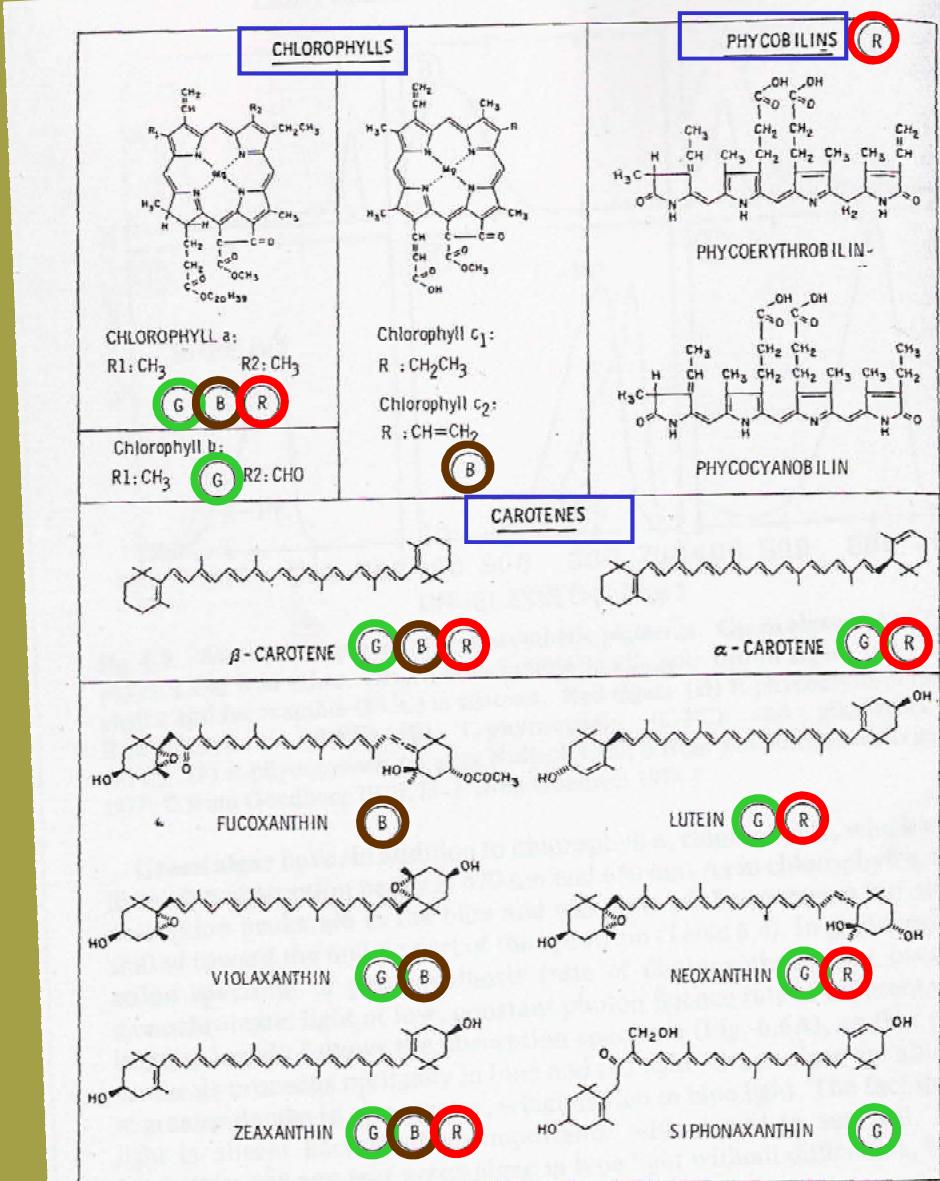


Fig. 6.4 Structural formulae of chlorophylls, chromophores of phycobilins, and carotenoids of seaweeds. G = green algae, B = brown algae, R = red algae. (Structural formulae from Ragan 1981.)

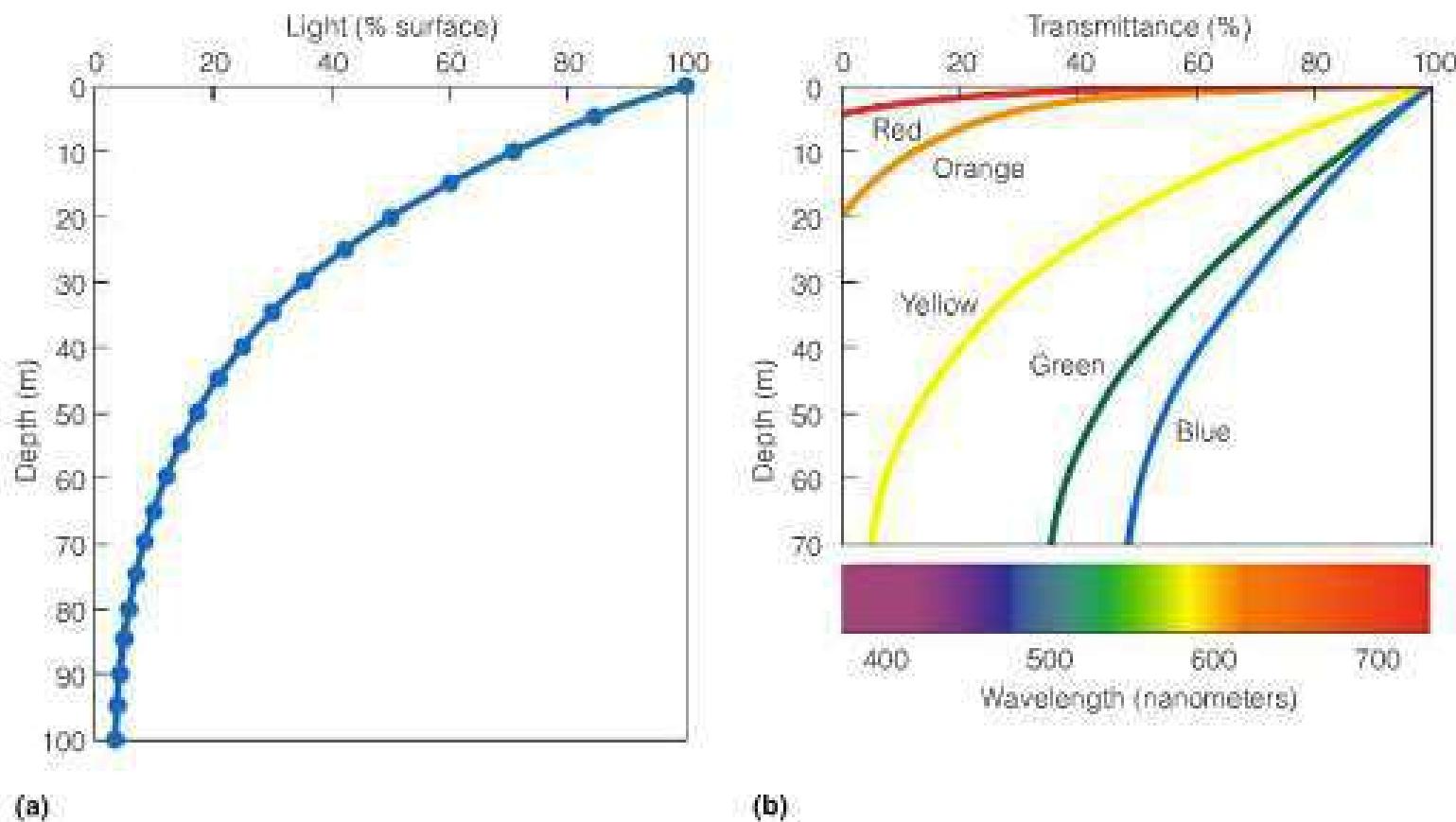


Figure 3.7 (a) Attenuation of incident light with water depth (pure water), expressed as a percentage of light at the water surface. Estimates assume a light extinction coefficient of $k_w = 0.035$ (see Quantifying Ecology 4.1, pp. 56–57). (b) The passage of light through water (transmittance) reduces the quantity of light and modifies its spectral distribution (see Figure 2.4). Red wavelengths are attenuated more rapidly than green and blue wavelengths.

Lo spettro di assorbimento della clorofilla a e b e di altri pigmenti

La maggior parte degli organismi fotosintetici usa le lunghezze d'onda nella banda blu-verde (450-500 nm) che è quella preferenzialmente trasmessa dall'acqua limpida. Molte alghe assorbono energia luminosa a lunghezze d'onda più lunghe

La finestra verde

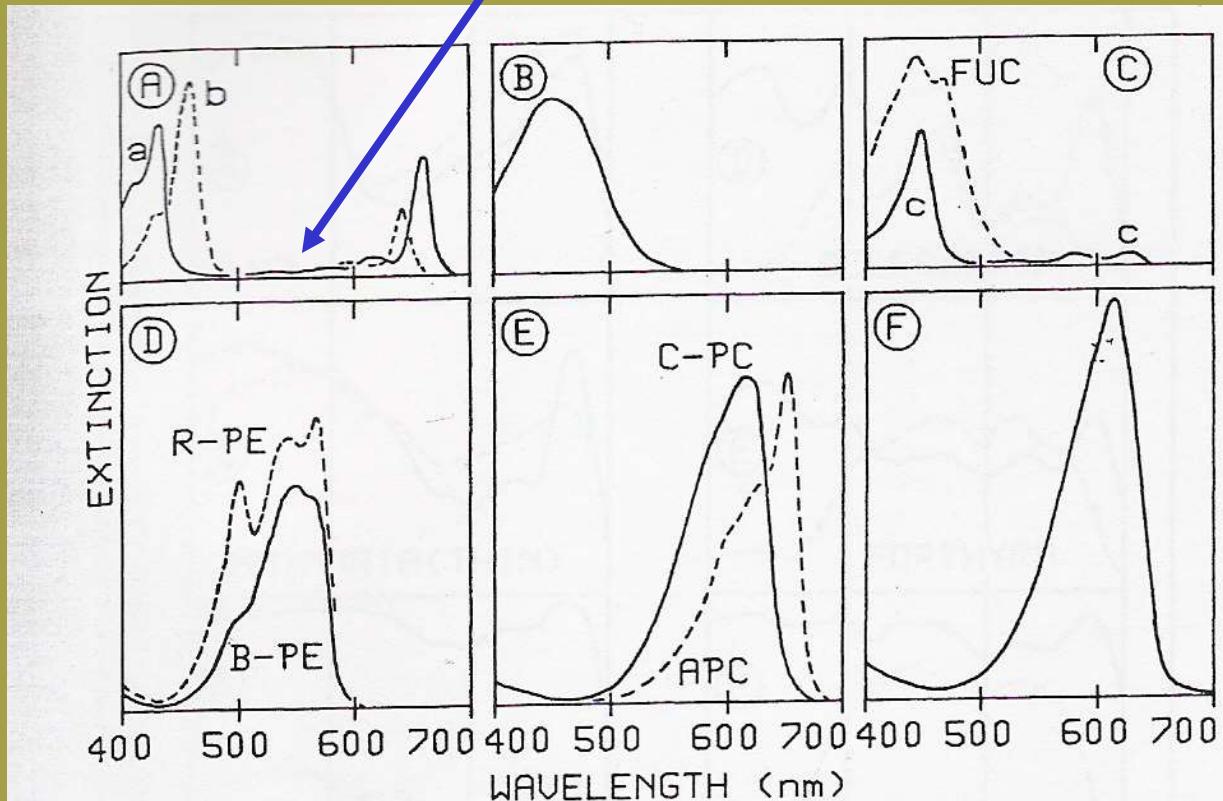


Fig. 6.5 Absorption spectra of photosynthetic pigments. **Green algae:** (A) Chlorophylls a and b in ether. (B) Siphonaxanthin in ethanol. **Brown algae:** (C) Chlorophyll c and fucoxanthin (FUC) in acetone. **Red algae:** (D) R-phycoerythrin (R-PE), B-phycoerythrin (B-PE). (E) C-phycocyanin (C-PC) and allophycocyanin (APC). (F) R-phycocyanin. (A after Nultsch 1986; B from Yokohama and Kageyama 1977; C from Goedheer 1970; D-F from Goodwin 1974.)

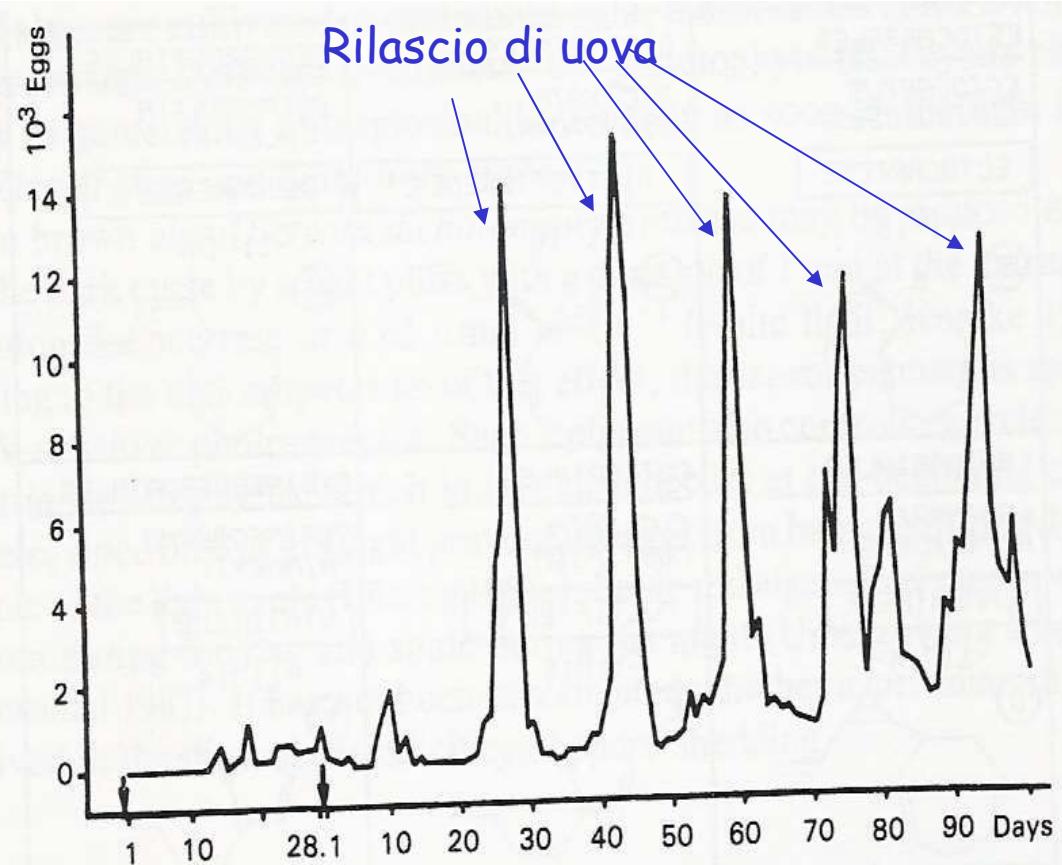


Fig. 6.16 Lunar periodism in the brown alga *Dictyota dichotoma*. Experimental female gametophytes were cultivated at 14 h of light per day. Number of eggs released was counted daily for more than three months. Continuous light was given on days 1 and 28 (arrows). Thereafter a semilunar rhythm of egg release was apparent. (From Müller 1962.)

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L'utilizzo

- Alimenti
- Estrazione di ficocolloidi
(medicinali, saponi, cosmesi)
Alginati
Agari
Carragenina
- Fertilizzanti per l' agricoltura
- Purificazione dell' acqua (Maerl)



Alimenti



Alimenti

KOMBU (Giappone)

HAIDAI (Cina)

WAKAME (Giappone)

NORI (Giappone)

Zicai (Cina)

Laminaria japonica, *Laminaria angustata*

Laminaria japonica

Undaria pinnatifida e altre

Porphyra tenera e altre

Porphyra haitanensis e altre

Estrazione di ficocolloidi (medicinali, saponi, cosmesi)

Alginati

Agari

Carragenina

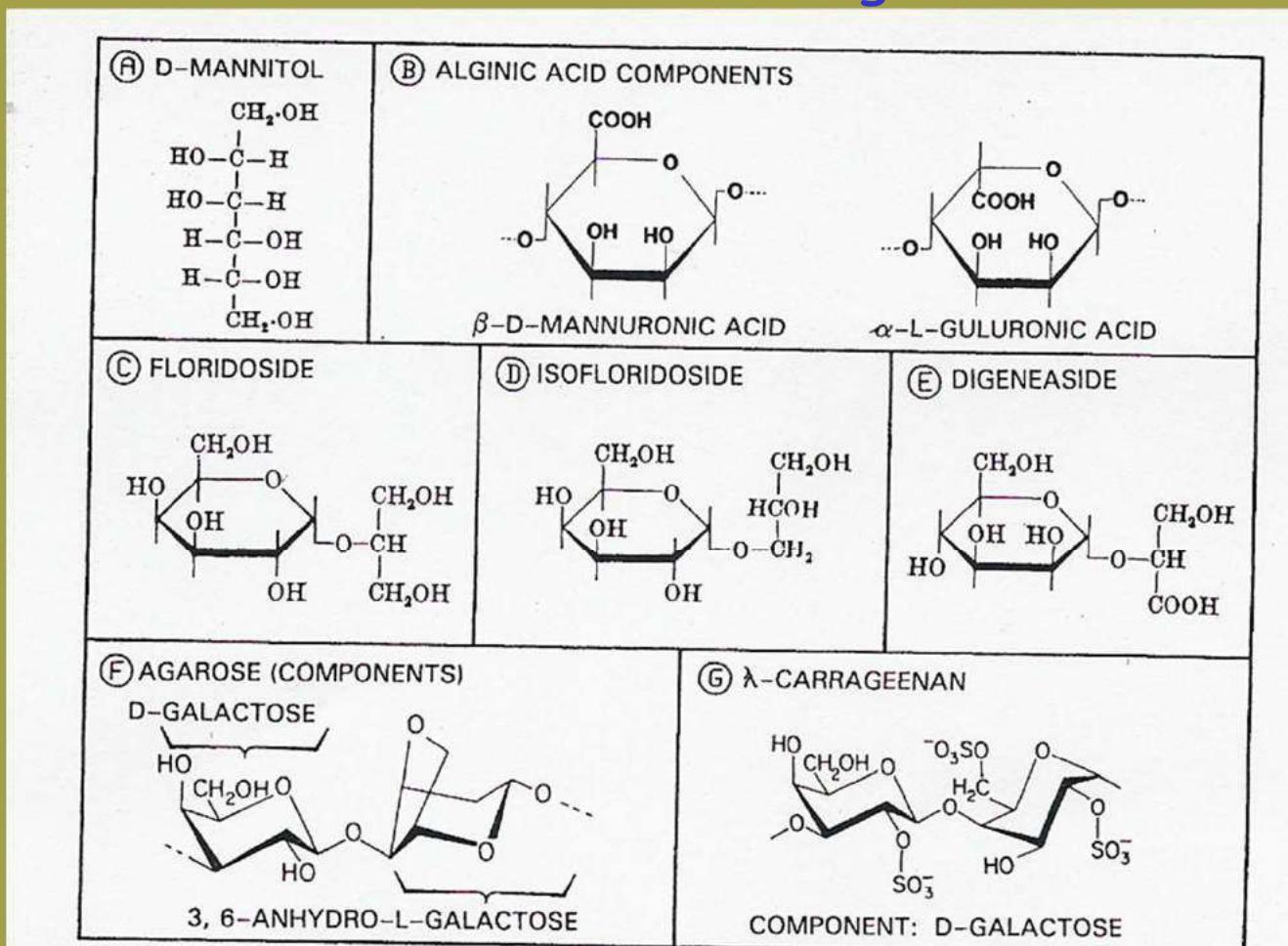


Fig. 8.7 Structural formulae of important seaweed carbohydrates. (Compiled after Percival 1979; Percival and McDowell 1967; McCandless 1981.)

AGAR

Gelidium, Pterocladia, Gracilaria

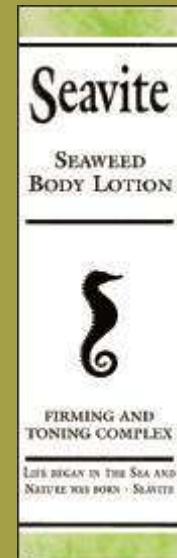
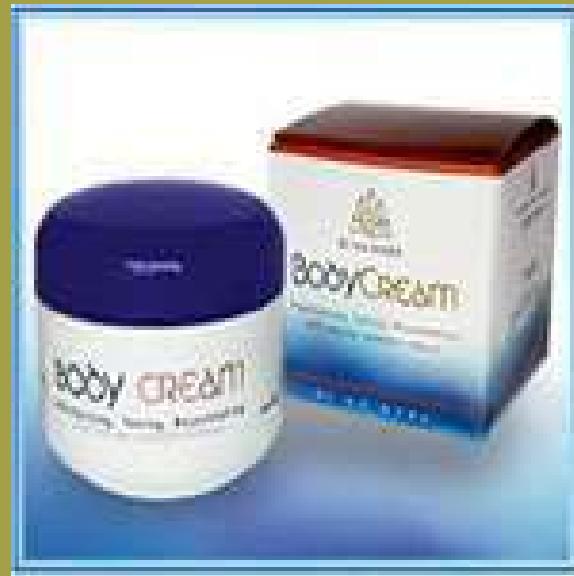
ALGINATI

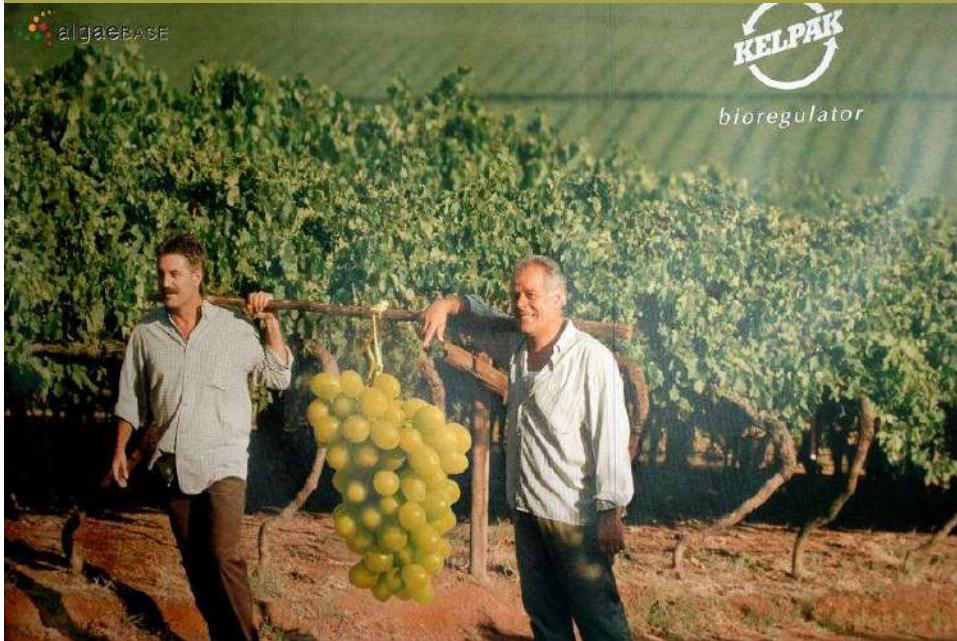
Laminaria, Macrocystis, Ascophyllum nodosum

CARRAGENINA

Chondrus, Mastocarpus

Cosmesi





Fertilizzanti liquidi



Fertilizzanti liquidi

Liquid extracts of marine brown algae are marketed for use in [agriculture and horticulture](#). Most of these extracts are prepared from dried *Ascophyllum nodosum* meal (e.g. "Maxicrop", manufactured in the United Kingdom), or from dried total drift, often referred to as "blackweed", but some utilise other species, such as *Fucus serratus* and *Laminaria species* (e.g. "SM3"; United Kingdom). One product currently being marketed is prepared from the stipes of *Ecklonia maxima* from South Africa (Kelpak, right). Other products prepared from seaweed include "Algifert" from Algea, a Norwegian company, "Seagro", manufactured in New Zealand; and "Seasol", an extract manufactured by a company in Tasmania. These are prepared from hot-water extracts of either the dried or wet seaweed, sometimes with the addition of sodium carbonate to aid extraction.

Liquid seaweed extracts are used at [very high dilution rates](#) which results in only very small quantities of material being applied to a given area. The active substances in the seaweed extracts must therefore be capable of having an effect at a low concentration.





algaebASE

Eklonia

Sud Africa











Ascophyllum nodosum, Irlanda











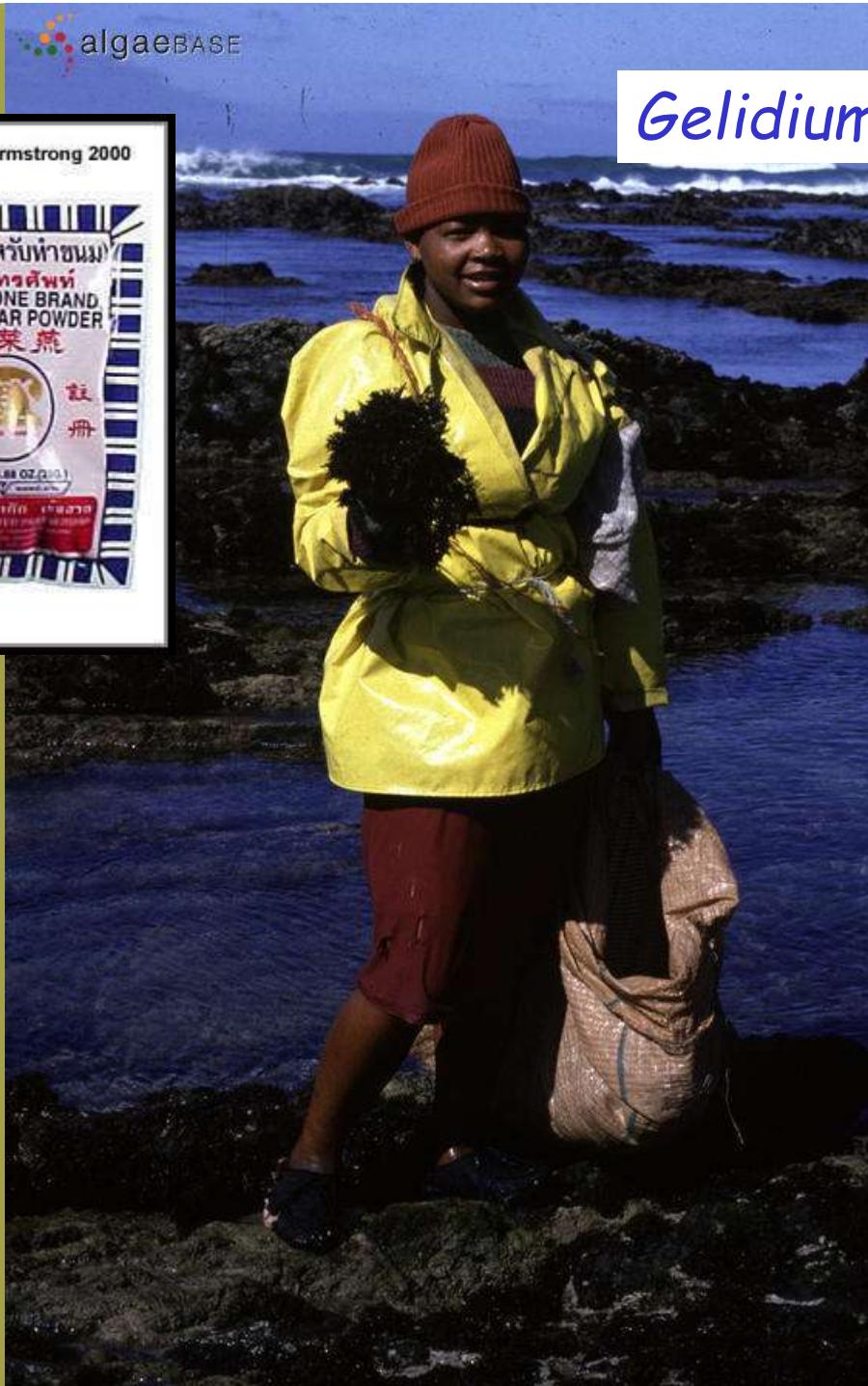
A. nodosum come cibo



Polvere di alginati



Gelidium, Sud Africa



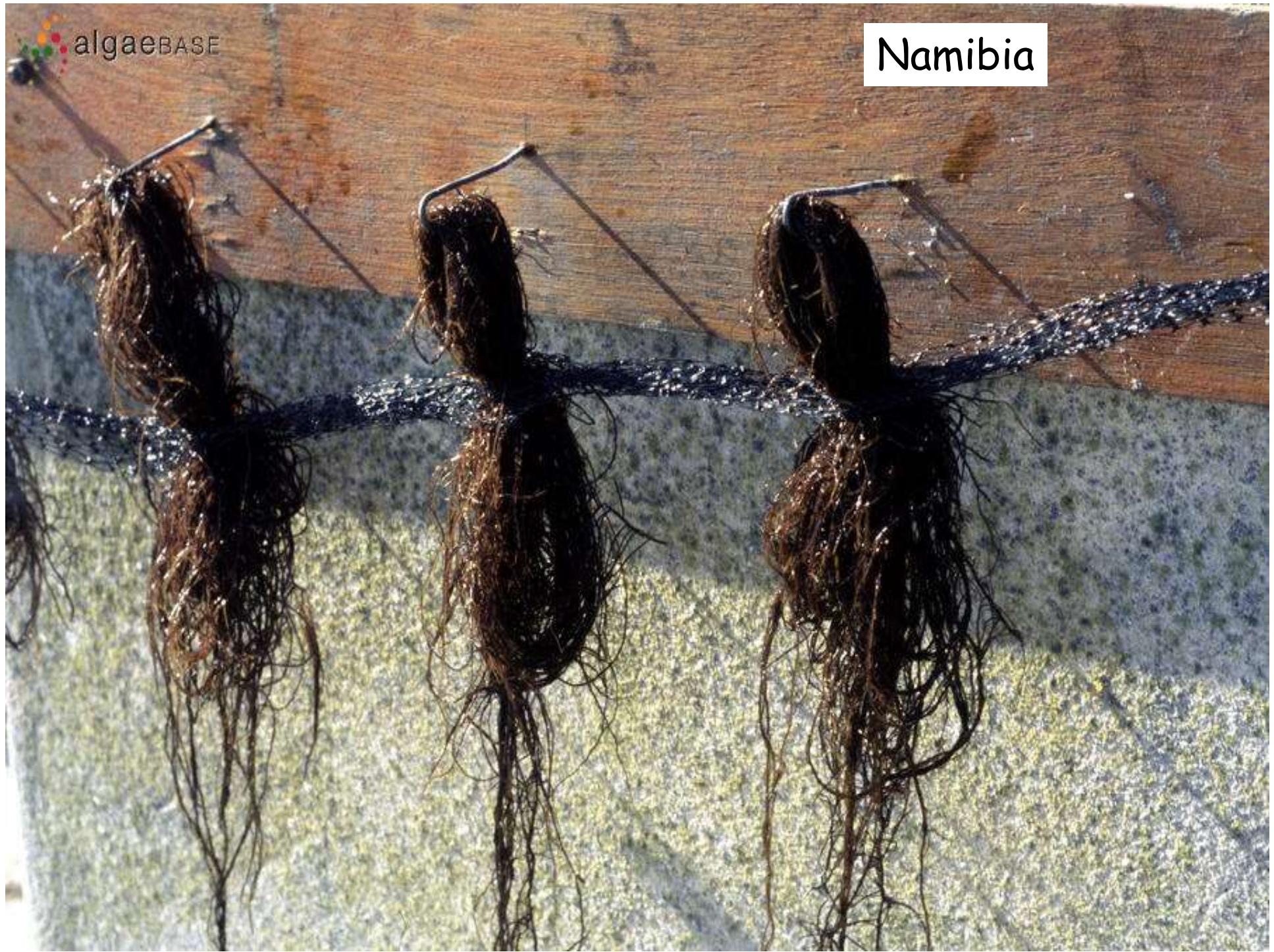
Agar agar

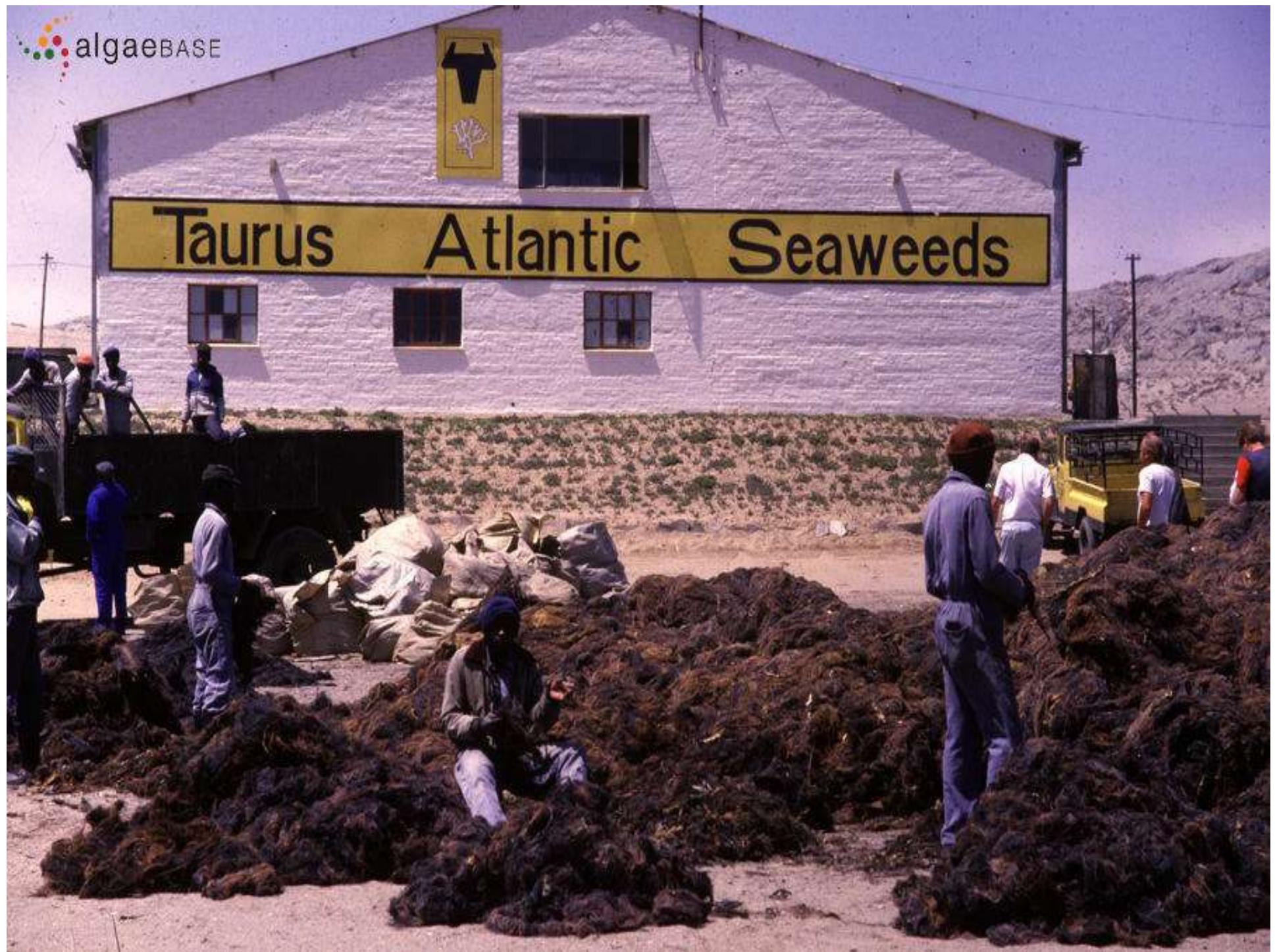


Gracilaria sp.



Namibia













Kappaphycus alvarezii (Floridophyceae)



K. alvarezii (purdoy)

K. alvarezii (tambalang green)



Bali, Filippine







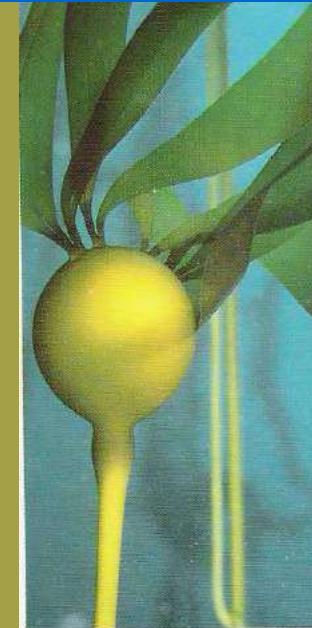






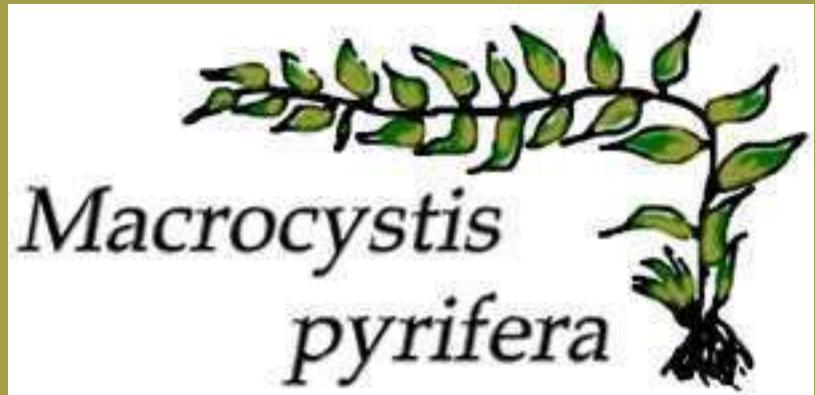
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Inflated like a balloon, this swollen bull kelp float holds a curious mixture of gases, including carbon monoxide.













Macrocystis pyrifera

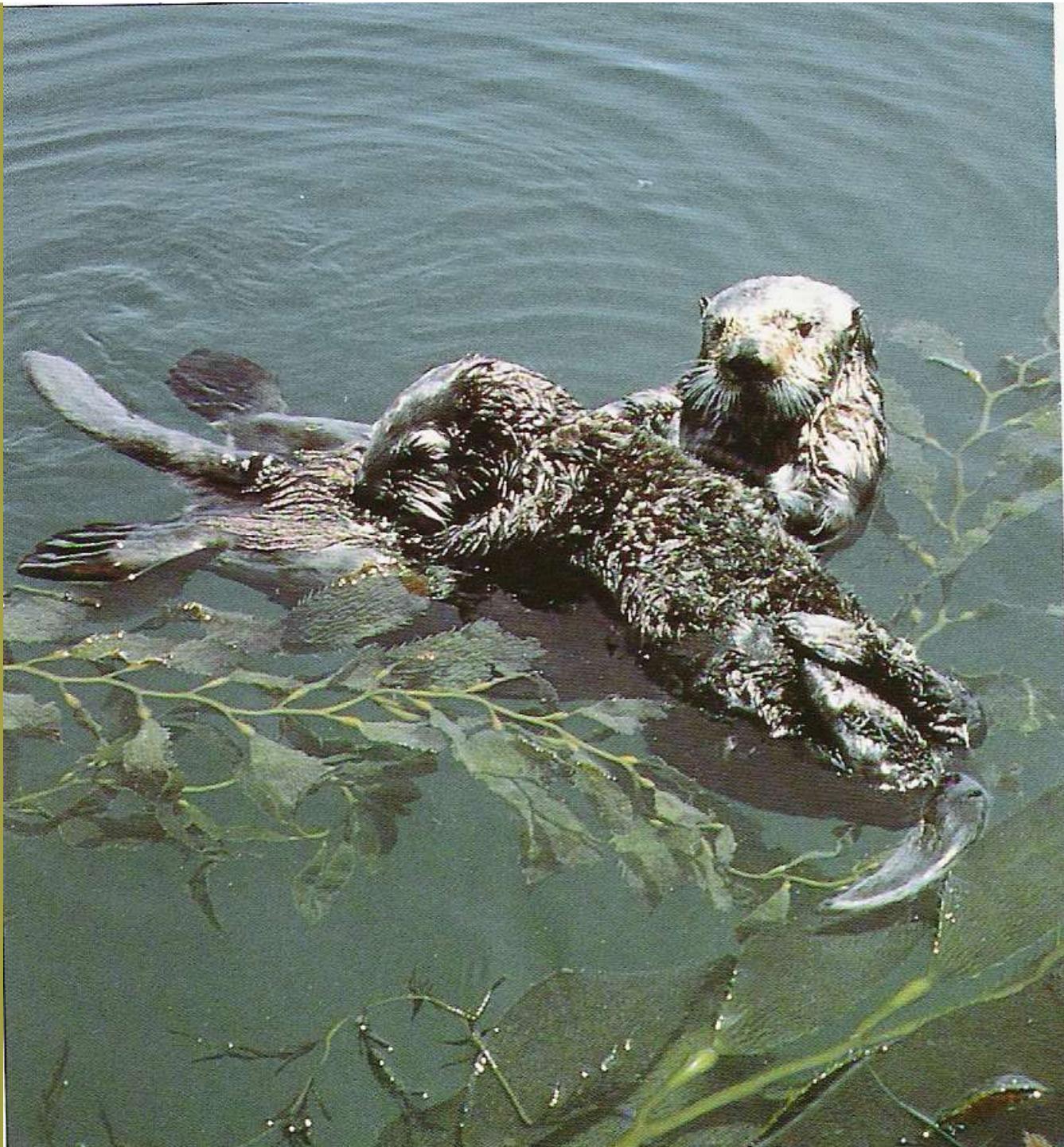












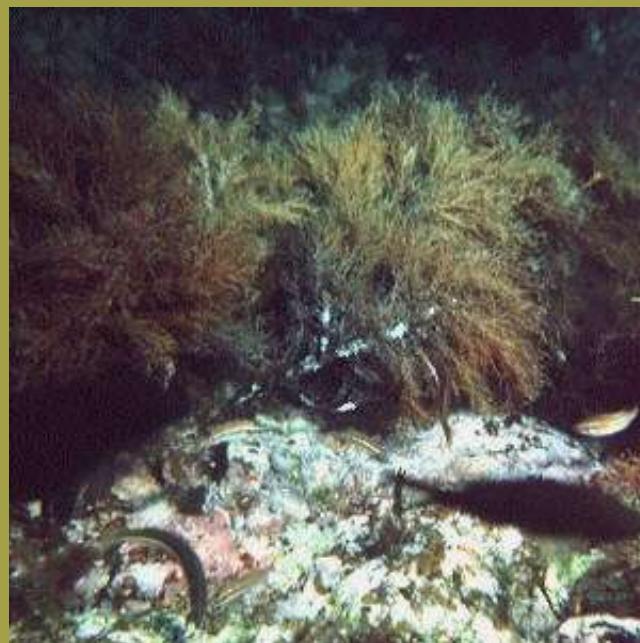












Cystoseira spp.

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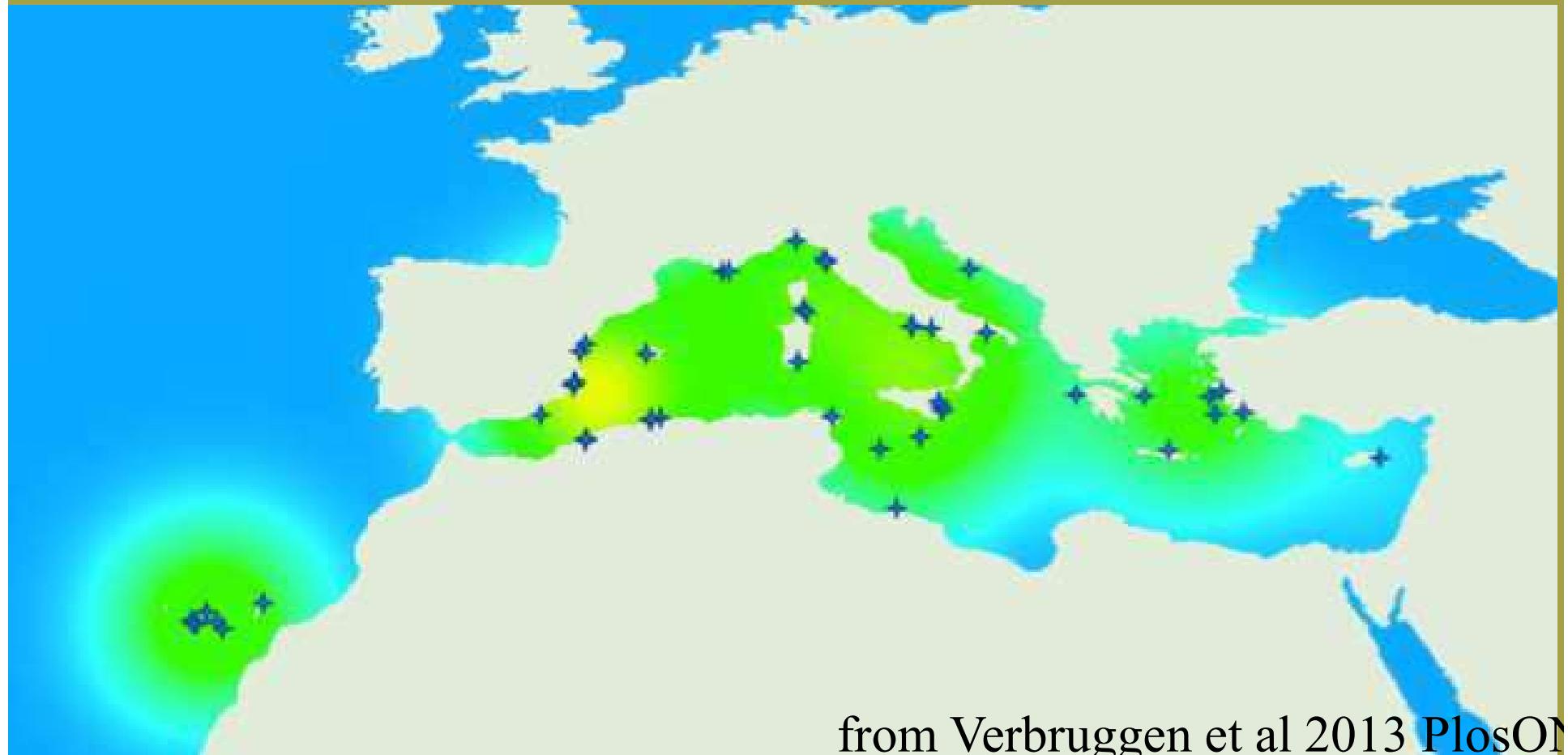
introduced species:

Macroalgae





Since the early 1990s it invades all kinds of substrata and depths.....



from Verbruggen et al 2013 PlosON

establishes dense monospecific stands, it easily overgrows and outcompetes seaweeds, all-sized) seagrass and invertebrates leading to an overall decrease in species diversity.

Mechanical destruction



Complexity of bottom

Canopy macrophytes

Algal Turfs

Eutrophication

Sedimentation

Mechanical destruction

Water movement

Other invaders

Herbivores

Other invaders



.....although introduced turf species advantage *C. cylindracea*

- Complexity of bottom
- Canopy macrophytes
- Algal Turfs
- Eutrophication
- Sedimentation
- Mechanical destruction
- Water movement
- Other invaders
- Herbivores

Herbivores



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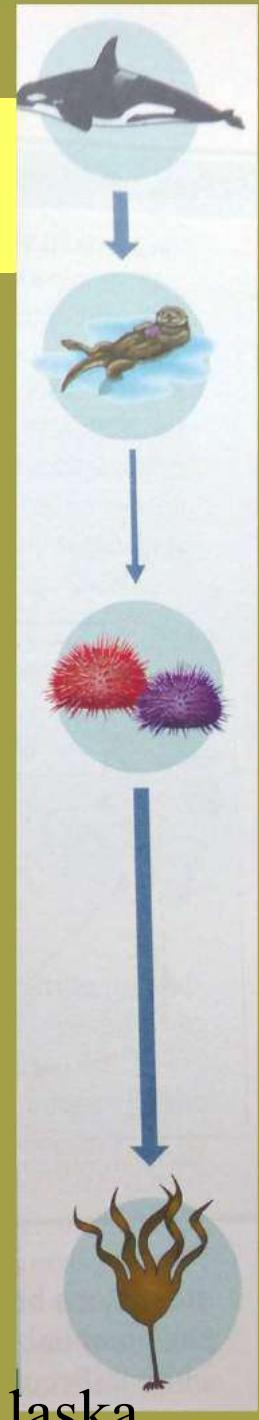
Global fishing of sea urchin predators and expanding sea urchin barrens

How are kelp forests stable?

Stability is due to top-down control effects

Test-crushing predatory fishes

Lobsters (*Panulirus interruptus* and *Jasus edwardsii*)





FEATURE ARTICLE: REVIEW

Sea urchin barrens as alternative stable states of collapsed kelp ecosystems

Karen Filbee-Dexter*, Robert E. Scheibling

Department of Biology, Dalhousie University, Halifax, Nova Scotia B3H 4J1, Canada

ABSTRACT: Sea urchin barrens are benthic communities on rocky subtidal reefs that are dominated by urchins and coralline algae; in the absence of intense herbivory by urchins, these barrens support luxuriant seaweed communities such as kelp beds (or forests). Barrens can extend over 1000s of km of coastline or occur in small patches (10s to 100s of m) within a kelp bed. They are characterized by low primary productivity and low food-web complexity relative to kelp communities and are generally considered a collapsed state of the kelp ecosystem. To assess the stability of sea urchin barrens and potential for return to a kelp-dominated state, we document temporal and spatial patterns of occurrence of barrens along temperate and polar coasts. We examine the various drivers of phase (or regime) shifts in these areas, the threshold levels of urchin abundance that trigger abrupt changes in ecosystem state, and the factors that maintain barrens once they have formed.



Sea urchins *Strongylocentrotus droebachiensis* graze along the edge of a kelp bed, creating a barren.

Photo: Robert Scheibling

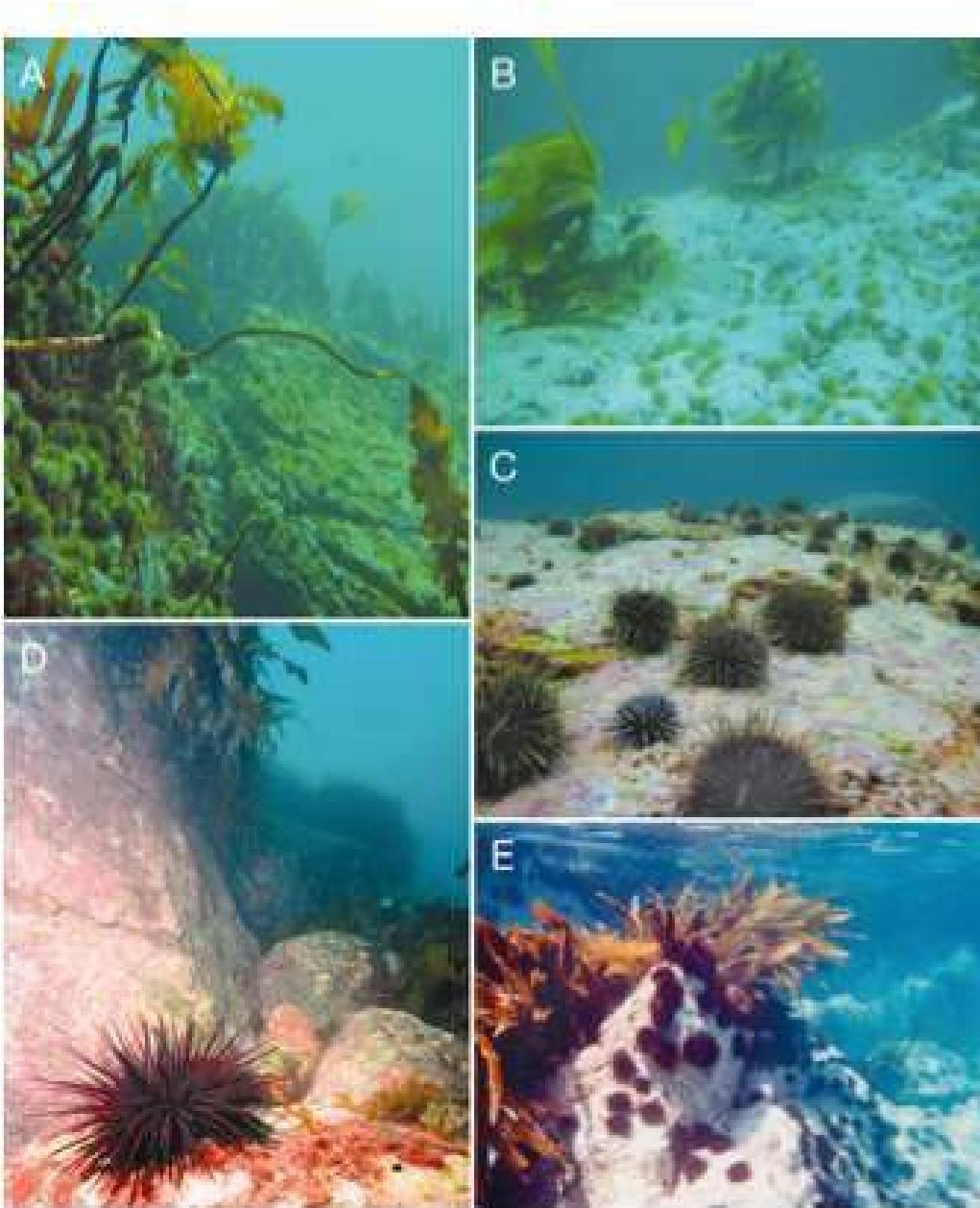
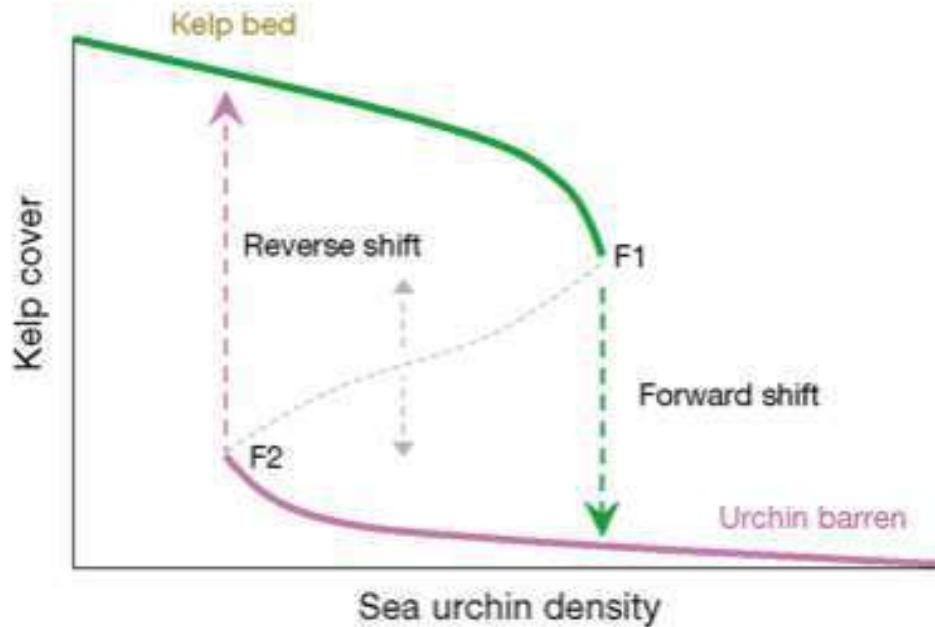
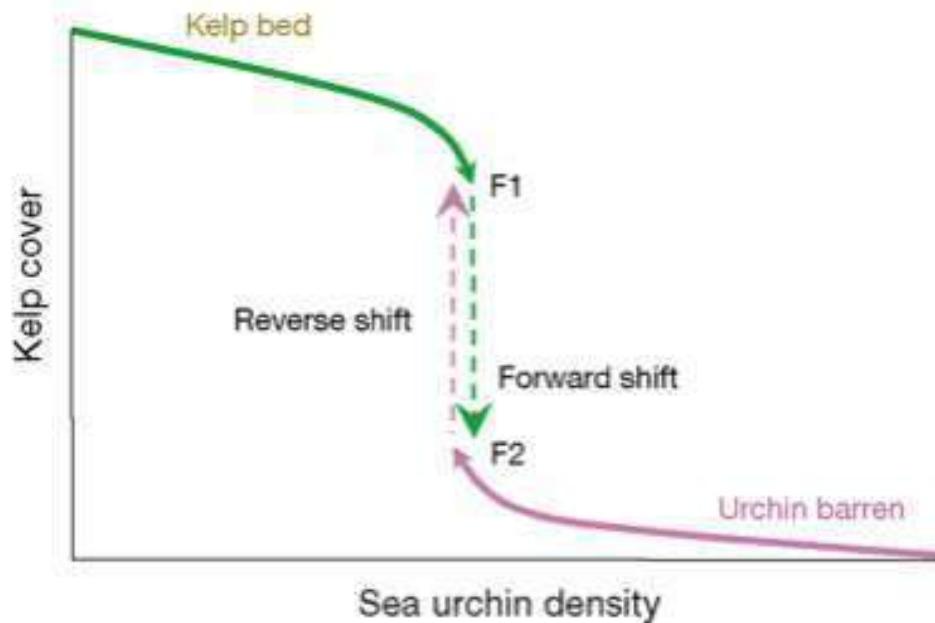


Fig. 1. (A) Destructive grazing front of sea urchins *Strongylocentrotus droebachiensis* advancing into a kelp bed near Halifax, Nova Scotia, Canada. Photo credit: R. E. Scheibling. (B) Extensive urchin (*S. polyacanthus*) barrens in the Aleutian Islands, USA. Photo credit: B. Konar. (C) Urchins *S. droebachiensis* on scoured coralline algae in barrens in Norway. Photo credit: C. W. Pageri. (D) Range-expanding urchin *Centrostephanus rodgersii* forming patchy barrens in a kelp bed in southeast Tasmania. Photo credit: S. D. Ling. (E) *S. nudus* grazing a kelp bed in Japan. Photo credit: D. Fujita

A. Discontinuous phase shift



B. Continuous phase shift



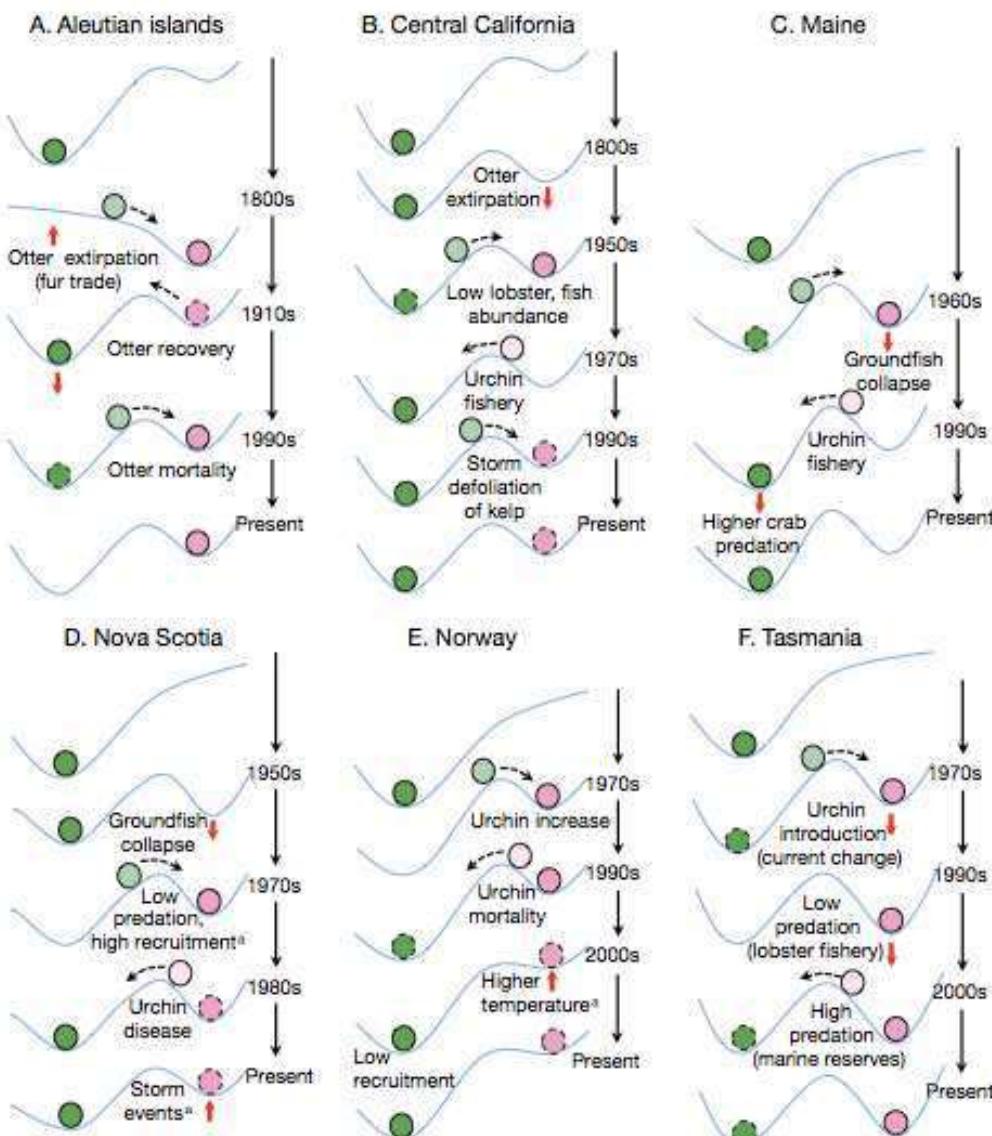
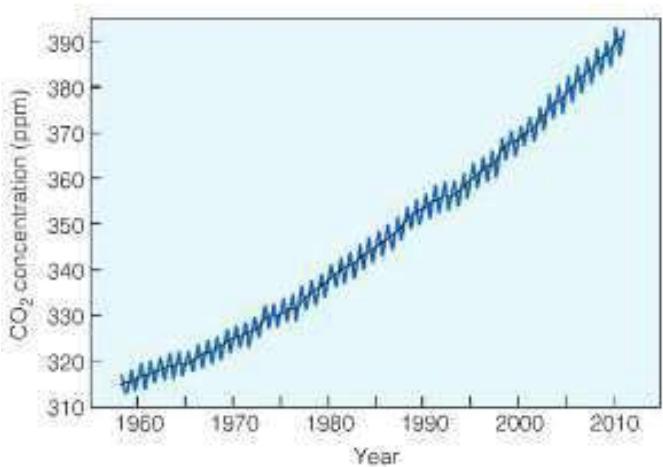
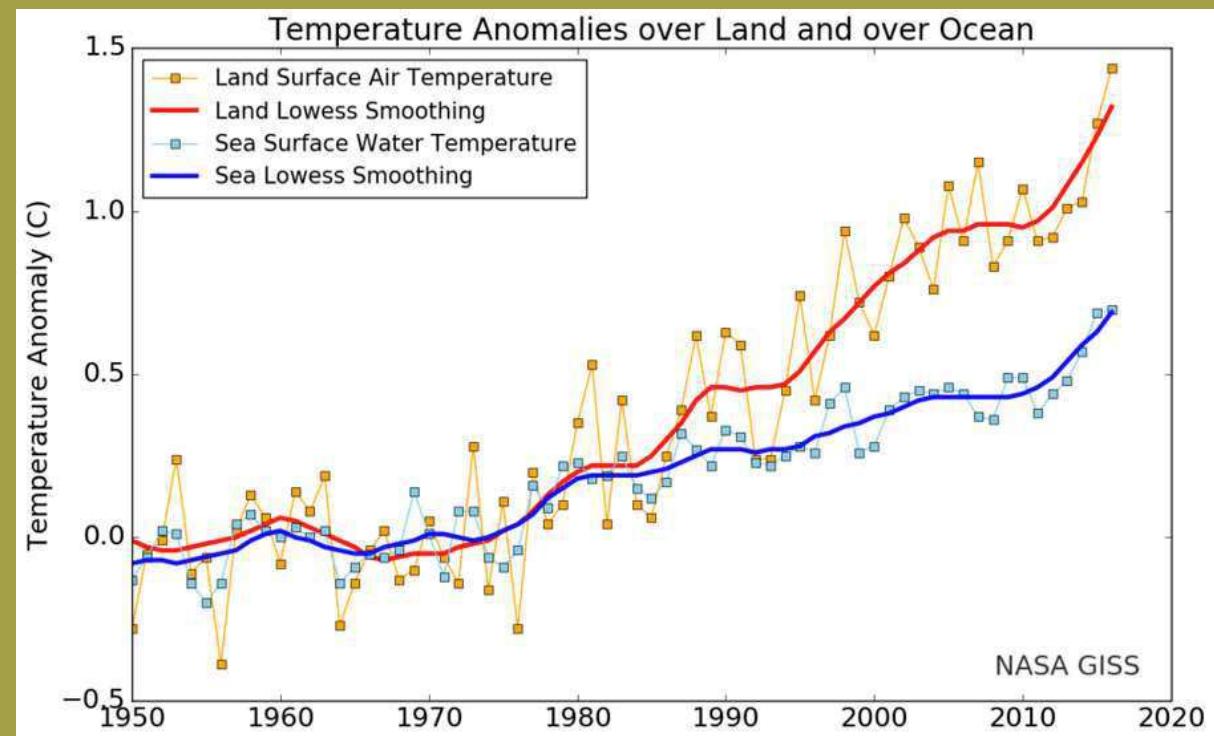


Fig. 5. Ball-in-cup diagrams representing phase shifts between kelp beds and sea urchin barrens in 6 regions: (A) Aleutian Islands, USA, (B) Central California, USA, (C) Maine, USA, (D) Nova Scotia, Canada, (E) Norway and (F) Tasmania, Australia. For each region, the top diagram (in chronological order) represents the earliest known community assemblage (determined by archeological evidence for the Aleutian Islands, California and Maine); this is followed by documented phase shifts and associated drivers leading to the present community state. See 'Barrens in regions with documented multiple phase shifts' for detailed explanation of drivers and dynamics for each region. Green balls represent kelp states, pink balls represent barren states and light green or light pink balls indicate a transitional stage (e.g. kelp bed with active urchin grazing patches or barrens with kelp regrowth). Balls with dashed lines represent patchy kelp or barrens; balls with solid lines represent extensive kelp or barrens. Red vertical arrows represent changes in domains of attraction (resilience); dashed black arrows represent shifts from one domain of attraction to another. ^aA statistical association, not a mechanistic driver

CO₂ in AIR

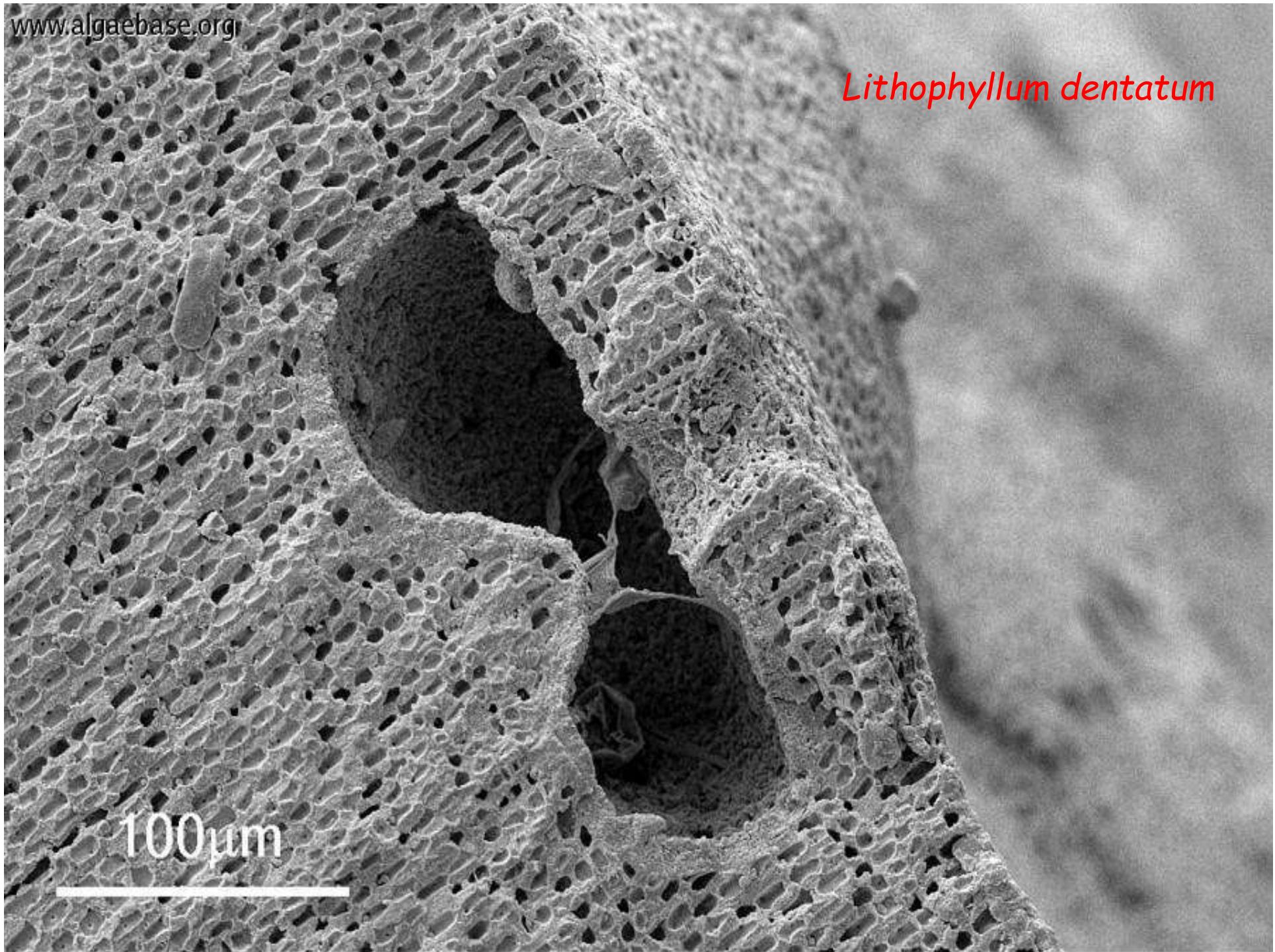


LAND and SEA temperature



Lithophyllum dentatum

100 μ m



sea acidification



Carbonic acid further dissociates into a hydrogen ion and a bicarbonate ion:

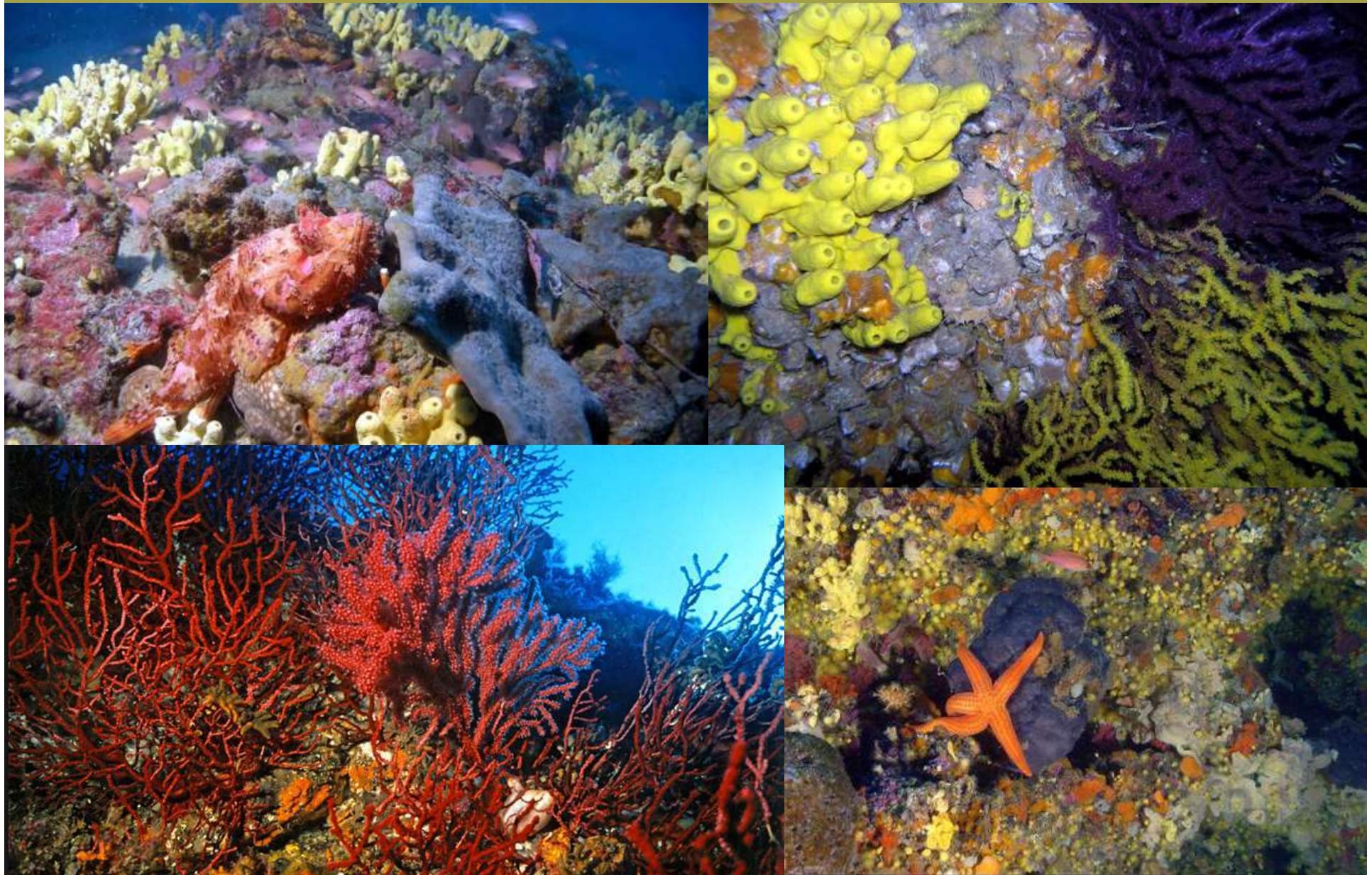


Bicarbonate may further dissociate into another hydrogen ion and a carbonate ion:

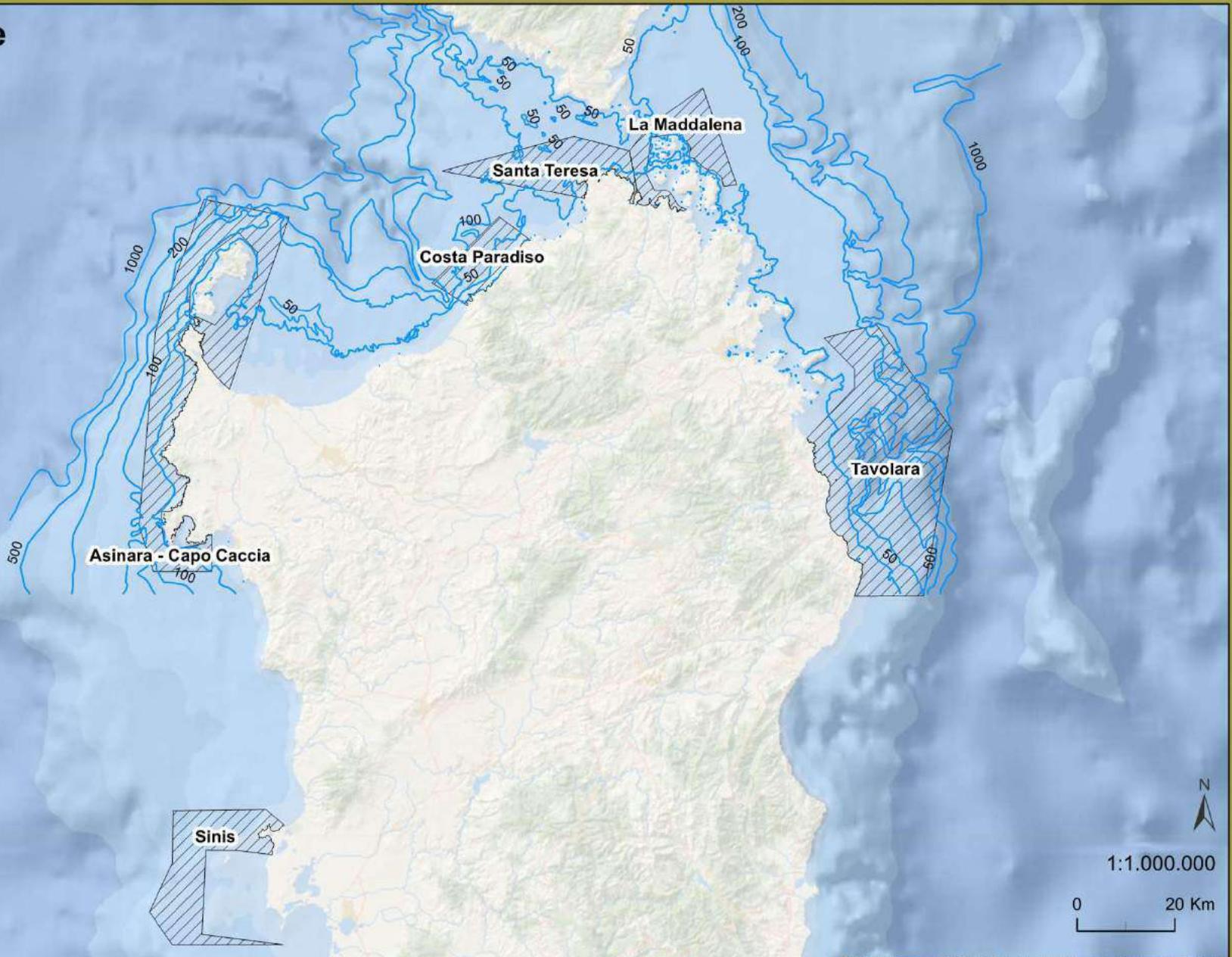


The carbon dioxide–carbonic acid–bicarbonate system is a complex chemical system that tends to stay in equilibrium.

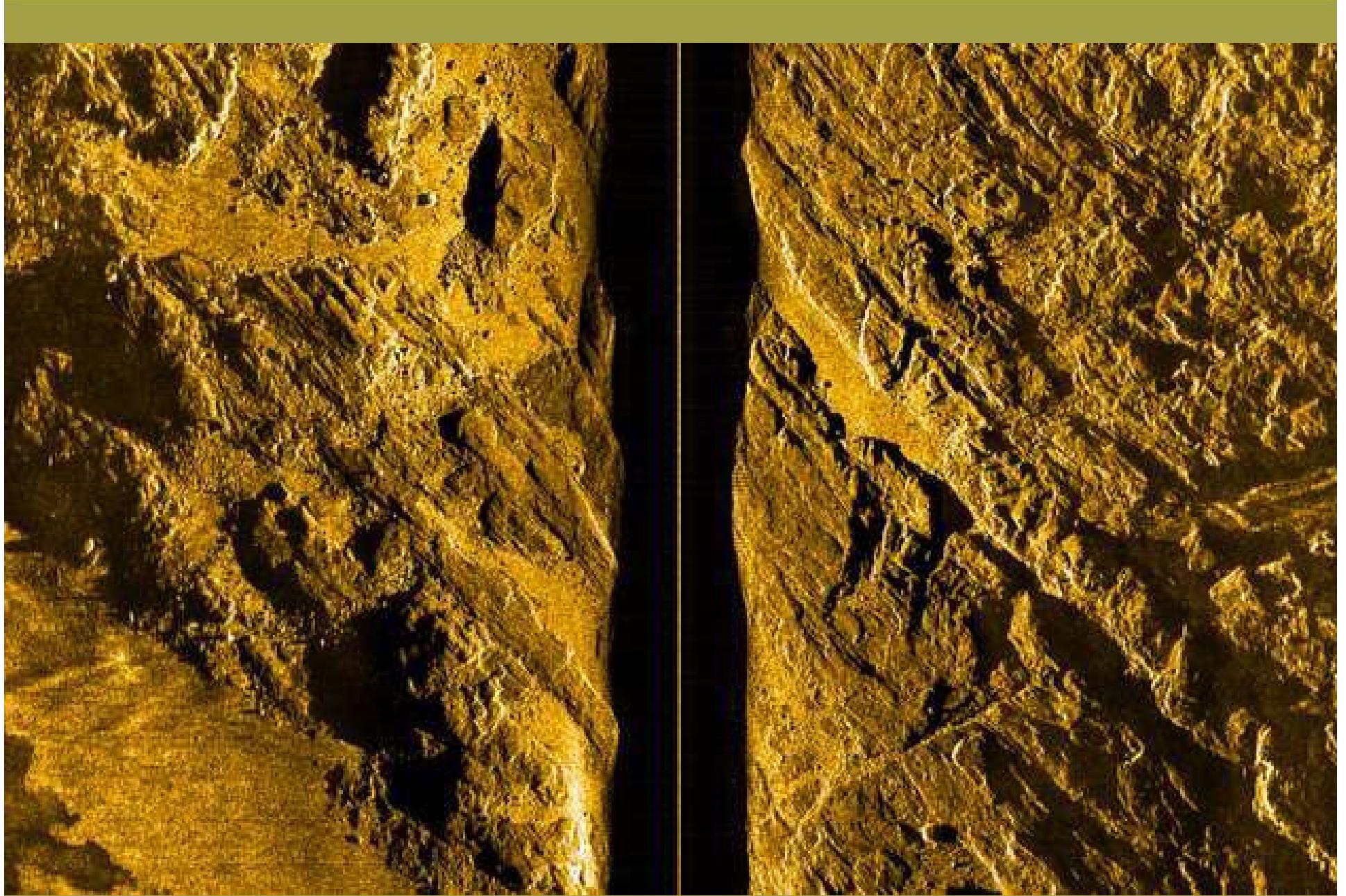
Mediterranean coralligenous

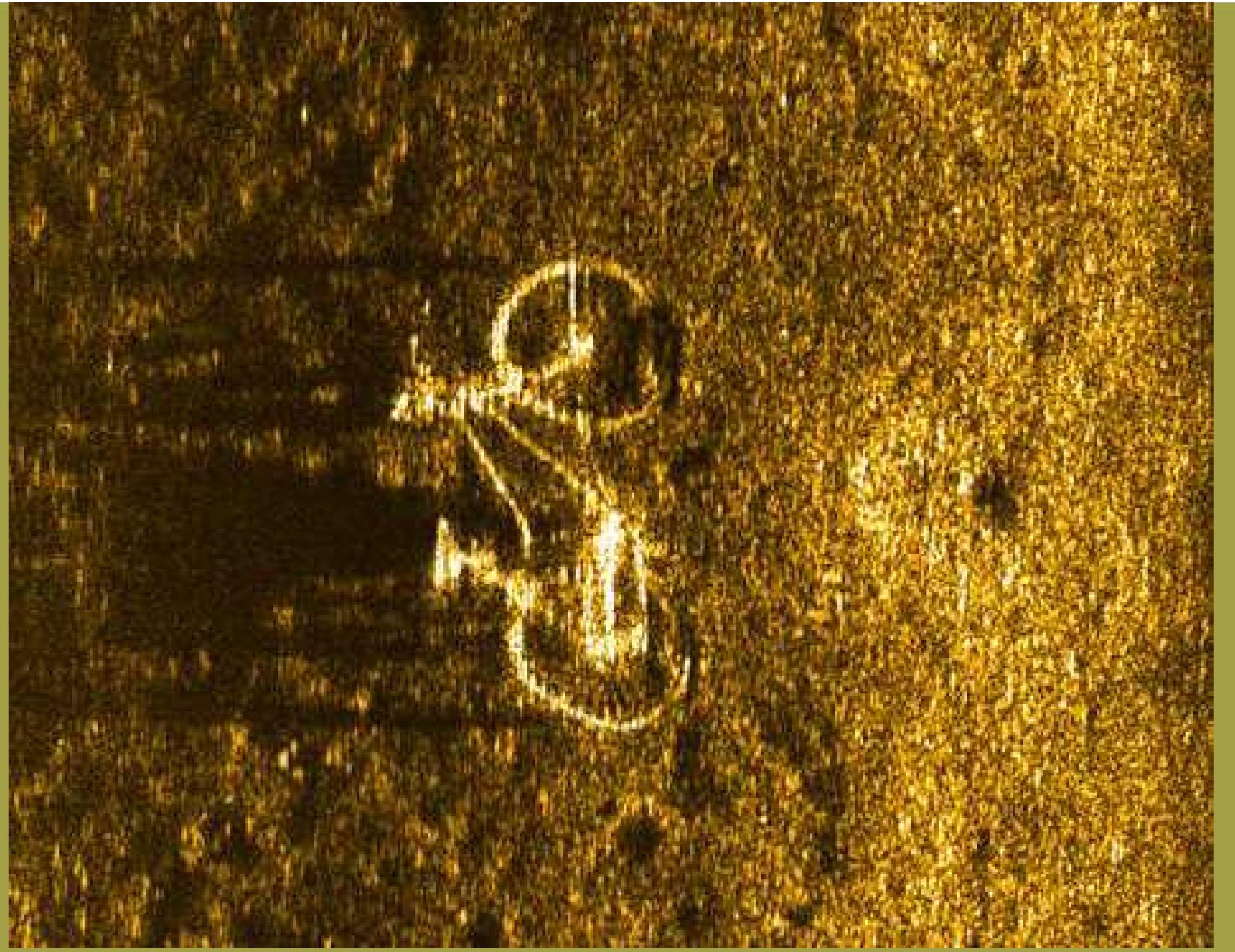


Macroaree











BRÖDAL